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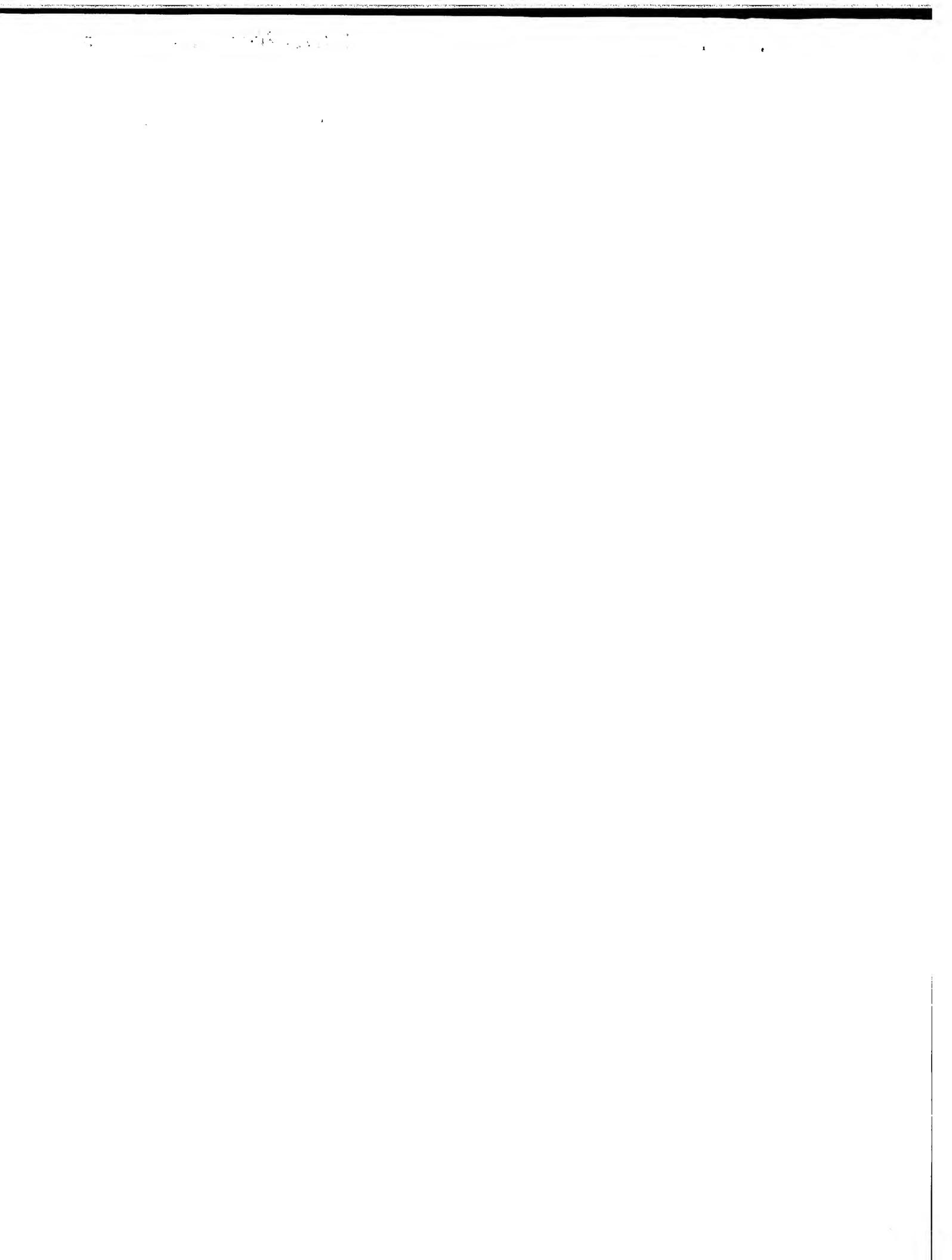
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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention:
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A vaccine composition comprising an immunoadjuvant compound consisting of a RHO GTPase family activator

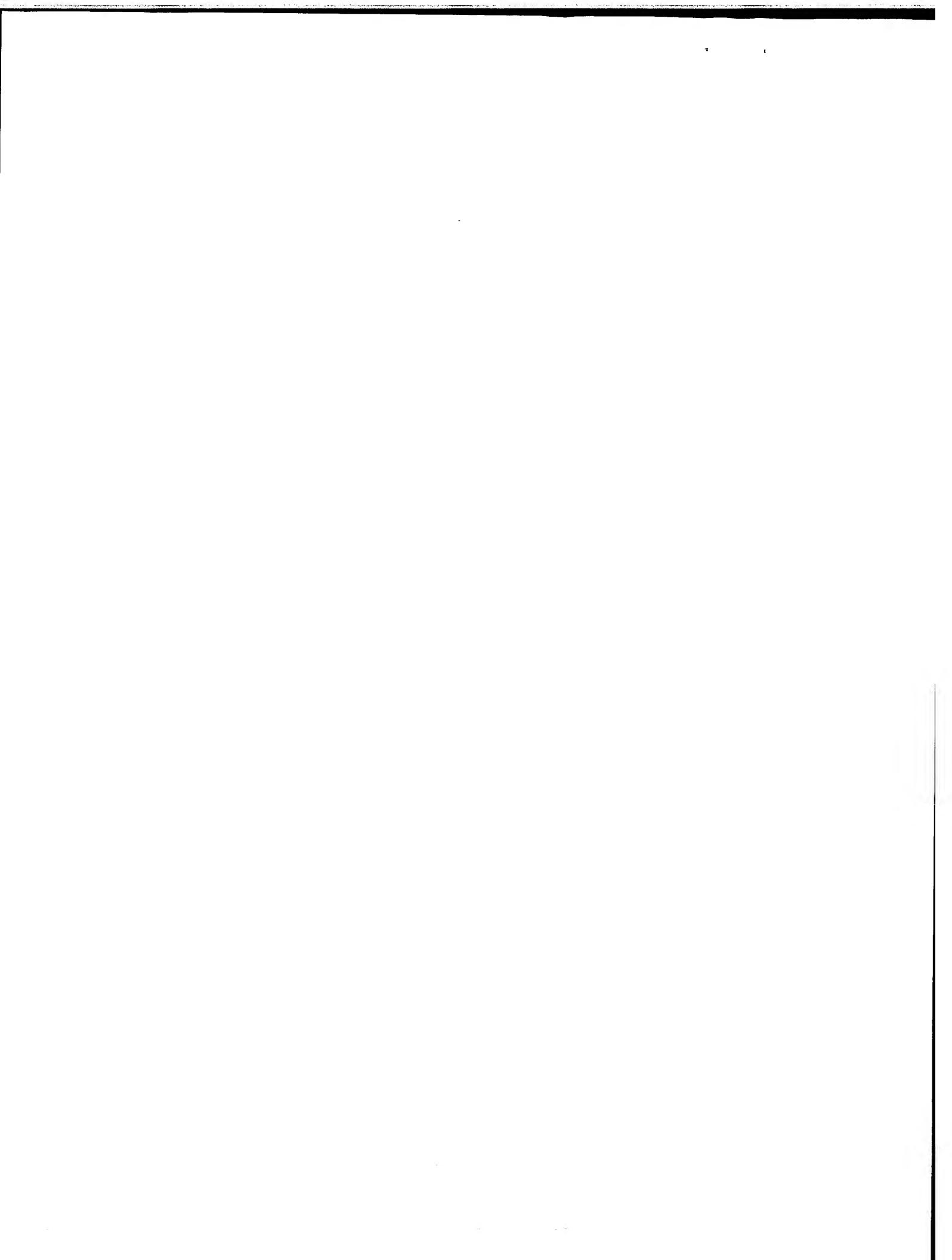
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Field of the invention

The present invention relates to a vaccine composition comprising an immuno adjuvant compound, wherein said immuno adjuvant compound consists of a RHO GTPase family activator.

Background of the invention

Vaccines have proven to be successful, highly acceptable methods for the prevention of infectious diseases. There are cost effective, and do not induce antibiotic resistance to the target pathogen or affect normal flora present in the host. In many cases, such as when inducing anti-viral immunity, vaccines can prevent a disease for which there are no viable curative or ameliorative treatments available.

Vaccines function by triggering the immune system to induce a response to an agent, or an antigen, typically in an infectious organism or a portion thereof that is introduced into the body in a non-infectious or non-pathogenic form.

Once the immune system has been "primed" or sensitised to the organism, later exposure of the immune system to this organism, results in a rapid and robust immune response that destroys the pathogen before it can multiply or infect enough cells in the host organism to cause disease symptoms.

The agent, or antigen, used to prime the immune system can be the entire organism in a less infectious state, known as an attenuated organism, or in some cases, component of the organism such as carbohydrate proteins or peptides representing various structural components of the organism.

In many cases, it is necessary to enhance the immune response to the antigens present in a vaccine in order to stimulate the immune system to a sufficient extent to make a vaccine effective, i.e., to confer immunity. Many proteins and most peptide and carbohydrate antigens, administered alone, do not elicit a sufficient antibody response to confer immunity. Such antigens need to be presented to the immune system in such a way that they will be recognized as foreign and will elicit an immune response.

To this end, additives like adjuvants, have been devised, which immobilise antigens and stimulate the immune response.

Recombinant proteins are promising vaccine or immunogenic composition candidates because they can be produced at high yield and 5 purity and manipulated to maximize desirable activities and minimize undesirable ones.

However, because they can be poorly immunogenic, methods to enhance the immune response to recombinant proteins are important in the development of vaccines or immunogenic compositions. Such 10 antigens, especially when recombinantly produced, may elicit a stronger response when administrated in conjunction with an adjuvant.

The best known adjuvant, Freund's complete adjuvant, consists of a mixture of mycobacteria in an oil/water emulsion.

Freund's adjuvant works in two ways; first, by enhancing cell and 15 humoral-mediated immunity, and second by blocking rapid dispersal of the antigens challenge, also called "depot effect". However, due to frequent toxic physiological and immunological reactions to this material, Freund's adjuvant cannot be used in humans.

Another molecule that has been shown to have stimulatory or 20 adjuvant activity is endotoxin, although known as lipopolysaccharide (LPS).

LPS stimulates the immune system by triggering an immediate 25 immune response, a response that has evolved to enable an organism to recognize endotoxin and the invading bacteria (of which it is a component) without the need for the organism to have been previously exposed. But LPS is although too toxic to be a viable adjuvant.

Thus, there is a recognized and permanent need in the art for new compounds which can be administered with antigens in order to stimulate the immune system and generate a more robust antibody response to 30 the antigen than will be seen if the antigens were injected alone.

Additionally, it should be noted that parenteral administration i.e. intramuscularly or sub-cutaneous, of antigens of vaccines are normally regarded as the most convenient way of administration.

However, the injection presents a range of disadvantages. It 35 requires the use of sterile syringes and may cause pains and irritations,

particularly in the case of repeated injections, including the risk of infection. More significantly, intramuscularly injections are often poorly tolerated. There is often likely to be indurations (hardening of tissue) haemorrhages and/or necrosis (local death of tissue) at the injection site.

5 Besides, untrained person cannot administer injections.

Based on these observations, it should be noted that mucosal immunity has take a considerable importance in vaccine development because nearly all viral, bacterial and parasitic agent that cause disease of the intestinal, respiratory and genital tracks enter through the mucosal barrier. Furthermore, mucosal and systemic immune responses are often elicited and regulated independently, and induction of protective immunity at the most frequent sites of entry is likely to be most effective. Additionally, young children and elderly individuals may respond better to mucosal vaccines because the mucosal immune system develops earlier and appears to remain functional longer than the systemic compartment. Mucosal immunisations are also easier and less expensive than systemic immunisations. For example, the existence of an oral polio vaccine has allowed immunisation campaigns that may soon eradicate polio worldwide.

20 Accordingly, it is also an object of the present invention to provide a vaccine composition comprising an immunoadjuvant compound which could be administered by the mucosal route. These and further objects will be apparent to one ordinary skill in the art.

25 **Summary of the invention**

The present invention is based on the experimental findings that an activator of Rho GTPases, namely the cytotoxic necrotizing factor 1 (cnf1) bears immunostimulatory properties towards the systemic and mucosal responses to orally administered ovalbumine, a prototype soluble protein antigen. CNF1 consists of an injection domain (amino acid residues 1-719 of SEQ ID N°1), allowing the binding and endosomal penetration of the toxin, followed by the intracytoplasmic injection of its catalytic domain (amino acid residues 720-1014 of SEQ ID N°1), responsible for Rho GTPases protein family activation.

A first object of the invention consists in a vaccine composition comprising an immunoadjuvant compound, wherein said immunoadjuvant compound consists of a Rho GTPase activator.

In another aspect, the invention relates to a vaccine composition
5 wherein said immunoadjuvant compound is selected from the group consisting of :

- a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°1,
- 10 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°2,
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°3,
- 15 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 1146 and ending at the amino acid residue 1451 of sequence SEQ ID N°4,
- a polypeptide comprising the amino acid sequence SEQ ID N°5,
- 20 - a polypeptide comprising the amino acid sequence SEQ ID N°6,
- a polypeptide comprising the amino acid sequence SEQ ID N°7,
- a polypeptide comprising the amino acid sequence SEQ ID N°8, and
- a polypeptide comprising the amino acid sequence SEQ ID N°9.

The present invention also relates to a vaccine composition wherein
25 the immunoadjuvant compound is a protein comprising a polypeptide consisting of; from the N-terminal end to the C-terminal end, respectively:

- a) the injection domain of a Rho GTPase activator , and
- b) the catalytic domain of a Rho GTPase activator.

30 **Description of drawings**

Figure 1: CNF1 effects on cell signaling pathway.

- 35 - 1A : Immunoblots showing the kinetics of CNF1-induced activation of Rho, Rac and Cdc42 in contrast to Ras, in HUVEC. Cells were

treated with 10^{-9} M CNF1 for different periods of time. Cell lysates were subjected to GST-fusion protein pull-down assays (noted GTPases-GTP). In parallel, 2% of each cell lysate were processed for immunoblotting to monitor their cellular depletion (noted Total-GTPases).

- 1B : Quantification of the CNF1-induced Rho protein activation. Immunoblots were scanned and quantified using N.I.H. Image 1.6. The level of activated Rho proteins was compared to the total Rho GTPase level present in 2% of control cell lysates (mean value of three independent experiments \pm SD).
- 1C : Immunoblots showing the interference of native CNF1 and catalytic inactive CNF1-C866S on cell signaling. HUVEC were treated with " 10^{-9} M" CNF1 or CNF1-C866S for the indicated periods of time, prior to immunoblotting analysis. MAP kinase signaling was investigated using anti-phosphop44/42 MAP Kinase (noted P-p44/42) and anti-phospho-p38 MAP Kinase (noted P-p38) antibodies. Jun kinase activity was investigated by anti-phospho-c-jun (noted P-c-jun) immunoblotting. NF-kappaB signaling pathway activation was investigated by following the I κ B α cellular depletion on immunoblots.

Figure 2 : Immunoadjuvant properties of orally administered CNF1

- 2A : Serum IgG antibody responses to orally co-administrated ovalbumin (OVA) and toxins. Five groups of mice were fed OVA alone or co-administrated with either CT (10 μ g), CNF1 (1 μ g or 10 μ g) or CNF1-C866S (10 μ g). Groups of eight mice were immunized with CNF1 or CNF1-C866S, whereas groups of four mice were immunized with OVA alone or CT. Groups of mice were challenged once, two weeks after the first immunization and sera collected 30 days after the first immunization. Data are expressed as geometric mean serum IgG anti-OVA Ab titers and individual titers are indicated (full circle). These results are representative of three independent experiments.

- 2B : IgG-subclasses distribution of serum anti-OVA antibody responses after oral immunization with OVA and native or mutated CNF1 (CNF1-C866S). Groups of 8 mice were challenged twice after the first immunization and sera collected 45 days after the first immunization. Levels of the anti-OVA Ig-subclasses are expressed as a geometric mean (histogram).
- 2C: Feeding CNF1 promotes intestinal antibody responses to co-administered OVA. Groups of 8 mice were immunized three times orally with OVA and either CNF1 or CNF1-C866S. Intestinal IgA antibody responses were determined by the PERFEXT method (Villavedra et al., 1997). Data are expressed as geometric mean IgA antibody titer (histogram).

15 **Figure 3 Measure of the immunoadjuvant properties and toxin activity of CNF1 and DNT.**

- 3A: Measure of the toxin activity of CNF1, CNF1-CTER (720-1014), DNT-CTER (1154-1451) estimated by HEp-2 cells multinucleation assay, as previously described (Lemichez et al., 1997). As previously reported, CNF1-CTER is poorly active on cells due to its inability to penetrate into the cytosol (Lemichez et al., 1997). DNT-CTER shows a one thousand lower activity, as compared to CNF1.
- 3B: Serum IgG antibody responses to orally co-administered ovalbumin (OVA) and DNT or CNF1-toxin catalytic domains. Groups of 4 mice were fed OVA alone or co-administered with either CNF1-CTER (720-1014) (100 μ g) or DNT-CTER (1154-1451) (100 μ g). For CNF1, a group of height mice were fed OVA and CNF1 (10 μ g). Mice were challenged once, two weeks after the first immunization and sera collected 30 days after the first immunization. Data are expressed as geometric mean serum IgG anti-OVA Ab titers.

DETAILED DESCRIPTION OF THE INVENTION

The inventors have found according to the invention that Rho GTPase activators bear immunoadjuvant properties *in vivo*, when co-administered with an antigen, like ovalbumin.

Rho proteins are essential regulatory molecules controlling the actin cytoskeleton organisation and dynamics to accomplish different tasks such as cell polarity, movement, differentiation and phagocytosis (Takai et al., 2001, Etienne-Manneville et al., 2002, Chimini and Chavrier, 10 (2000)). Importance of Rho proteins in physiology is also evidenced by their direct or indirect implication as part of signaling molecules found mutated in human genetic disorders, as well as targets of numerous bacterial virulence factors and toxins (Boettner and Van Aelst, (2002) Boquet and Lemichez, (2003).

15 Rho proteins interfere with a large variety of signaling pathways controlling gene transcription (Bishop et al., 2000). Among them, a recent report has evidenced the activation of Rac and Cdc42 downstream the Toll-like receptor 2, a gram positive pathogen molecular pattern recognition receptor (PAMP) (Arbibe et al. (2000), Medzhitov et al. 20 (2002).

Also exemplifying the inter-relation between Rho proteins and the host defences is the Rac, Cdc42, VAV and WASP formation of a supramolecular activation complex (SMAC or "immunological synapse" crucial for lymphocyte activation (Krawczyk et al. 2001).

25 Many different pathogenic bacteria have evolved virulence factors and toxins aimed at mimicking an activation of Rho GTPase protein family, naturally occurring in eukaryotic cells via specific regulators namely GEF (for guanine nucleotide exchange factors). These cellular GEF consist in domains comprised in large proteins as best described for 30 Dbl (Olson et al., 1996; Schmidt and Hall 2002). Despite their lack of sequence homologies, virulence factors of pathogenic bacteria, for instance SopE and SopE2 from *Salmonella* have a GEF-like activity (Galan et al., 2000). Some other known factors of pathogenic bacteria, namely IpaC from *Shigella* and CagA from *Helicobacter*, activate Rho 35 GTPases by yet uncharacterised molecular mechanisms (Tran Van

Nhieu et al., 2000; Boquet and Lemichez 2003). Finally, a group of bacterial toxins comprising CNF1 also activates Rho proteins through a post-traductional modification (Boquet and Lemichez 2003)

According to the invention, the inventors have now surprisingly
5 found that the cytotoxic necrotising factor 1 (CNF1), has immunoadjuvant properties. More precisely, the inventors have found that CNF1 bears immunostimulatory properties toward the systemic and mucosal responses to orally administrated ovalbumin in mice.

Additionally, the inventors have found that a mutant of CNF1,
10 namely CNF1-C866S, a catalytically inactive mutant of CNF1 toward GTPases, in contrast to the wild type toxin, does not stimulate the systemic and mucosal responses to ovalbumin. This result points for Rho GTPases proteins activation being directly involved in the immunostimulatory effects of CNF1.

15 Supporting this point, the inventors have also found according to the invention that the catalytic domain of CNF1, and the catalytic domain of DNT, another Rho GTPase activator, bear also immunoadjuvant properties *in vivo*, when co-administered with an antigen, like ovalbumin.

Taken together, these results demonstrate clearly that different
20 Rho GTPases activators, structurally different, have immunoadjuvant properties.

Furthermore, the inventors have found that non neutralizing anti-
25 CNF1 antibodies are naturally found in humans, and that CNF1 activates the Rho GTPase proteins only transiently. Taken together these results demonstrate that CNF1 can be used as an immunoadjuvant compound, deserved of adverse effects such as the toxic effects described for LPS or Cholera Toxin B.

Accordingly, a first object of the invention consists in a vaccine composition comprising an immunoadjuvant compound, wherein said immunoadjuvant compound consists of a Rho.GTPase activator.
30

By "immunoadjuvant" it is herein intended a substance enhancing the immunogenicity of an antigen. By "Rho GTPase activator" it is intended herein a compound, which maintains Rho GTPases in a form bound to GTP. By "Rho GTPases", the one skilled in the art will understand the proteins belonging to the Rho GTPase family, which
35

encompasses RhoA, RhoB, RhoC, Rac1, Rac2 and Cdc42. (Burridge and Wennerberg, 2004).

5 The level of Rho GTPase bound to GTP can be easily measured by the methods, referred by those skilled in the art as GST-pull down assays and described for RhoA, B and C by Ren et al., 1999 and for Rac1, Rac2 and Cdc42 by Manser et al., 1998. These methods are described in the section Materials and methods.

10 The invention also concerns a vaccine composition as described below, wherein said immunoadjuvant is selected from the group consisting of :

- 15 a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°1,
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°2,
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°3,
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 1146 and ending at the amino acid residue 1451 of sequence SEQ ID N°4,
- a polypeptide comprising the amino acid sequence SEQ ID N°5,
- a polypeptide comprising the amino acid sequence SEQ ID N°6,
- a polypeptide comprising the amino acid sequence SEQ ID N°7,
- a polypeptide comprising the amino acid sequence SEQ ID N°8, and
- a polypeptide comprising the amino acid sequence SEQ ID N°9.

20 A Rho GTPase activator encompasses peptides comprising the amino acid sequence of interest starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°1 described above, and comprising a N-terminal amino acid sequence, linked to the amino group of the residue 720 of sequence SEQ ID N°1.

25 Preferably, the N-terminal amino acid sequence has a length up to 800 amino acid residues.

Preferably, the N-terminal amino acid sequence is homologous to a part or to the full length amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 719 of CNF1 of SEQ ID N°1.

5 In such a case, the N-terminal amino acid sequence can comprise substitutions of non-essential amino acid comprised in the sequence starting at the amino acid residue 1 and ending at the amino acid residue 719 of CNF1 of SEQ ID N°1.

10 A "non essential" amino acid residue is an amino acid residue that can be altered from the wild type sequence of CNF1 without altering the activating properties of Rho GTPases, whereas an "essential" amino acid residue is required for biological activity.

15 A Rho GTPase activator encompasses also peptides comprising two or more repeated motifs of the sequence 720-1014 of interest. In such a case, said peptide can comprise also an N-Terminal sequence as defined above.

20 A Rho GTPase activator encompasses also peptides structurally similar to those described above, derived from the catalytic domain of CNF2 of sequence SEQ ID N°2, the catalytic domain of CNF_Y of sequence SEQ ID N°3 and the catalytic domain of DNT of sequence SEQ ID N°4.

25 The use of the catalytic domain of Rho GTPase activator, as described above, is of particular interest. Indeed, as demonstrated in example 3, in the case of CNF1, and DNT, the use of the catalytic domain of these proteins is less toxic for cells than the overall proteins, but is sufficient to confer immunoadjuvanticity.

A Rho GTPase activator encompasses also peptides comprising :

- the amino acid sequence SEQ ID N°5 corresponding to SOPE, or
- the amino acid sequence SEQ ID N°6 corresponding to SOPE2, or
- 30 - The amino acid sequence SEQ ID N°7 corresponding to IpaC, or
- the amino acid sequence SEQ ID N°8 corresponding to CagA, or
- the amino acid sequence SEQ ID N°9 corresponding to the GEF sequence of Dbl,

35 which include more amino acids, and exhibit at least the same activity towards Rho GTPase activation.

Alternatively, the immunoadjuvant according to the invention is selected from the group consisting of :

- a polypeptide comprising the amino acid sequence SEQ ID N°1,
- a polypeptide comprising the amino acid sequence SEQ ID N°2,
- 5 - a polypeptide comprising the amino acid sequence SEQ ID N°3, and
- a polypeptide comprising the amino acid sequence SEQ ID N°4.

Another object of the invention consists in a vaccine composition, wherein said immunoadjuvant compound is a protein comprising a polypeptide consisting of; from the N-terminal end to the C-terminal end, 10 respectively:

- a) the injection domain of a Rho GTPase activator , and
- b) the catalytic domain of a Rho GTPase activator.

By "injection domain of a Rho GTPase activator" it is intended herein, an amino acid sequence allowing the binding and intracellular penetration of a catalytic domain of a Rho GTPase activator. 15

By "catalytic domain of a Rho GTPase activator" it is intended herein, an amino acid sequence able to activate a Rho GTPase.

The attachment of the injection domain to the catalytic domain above mentioned, to produce a fusion protein may be effected by any 20 means which produces a link between the two constituents, which is sufficiently stable to withstand the conditions used and which does not alter the function of either constituent.

Preferably, the link between them is covalent.

Numerous chemical cross-linking methods are known and 25 potentially applicable for producing the fusion protein. For example, non-specific chemical cross-linking methods, or preferably methods of direct chemical coupling to a functional group, found only once or a few times in one or both of the polypeptides to be cross-linked.

Coupling of the two constituents can also be accomplished via a 30 coupling or conjugating agent. There are several intermolecular cross-linking reagents, which can be used (see, for example, Means, G. E. et al. (1974)). Among these reagents are, for example, N-succinimidyl 3-(2-pyridyldithio) propionate (SPDP) or N, N'-(1,3-phenylene) bismaleimide.

Cross-linking reagents may be homobifunctional, i.e., having two functional groups that undergo the same reaction such as bismaleimidohexane ("BMH").

Alternatively, to solve the problems of protein denaturation and 5 contamination during chemical conjugation, recombinant techniques can be used to covalently attach the polypeptide of interest to the virulence factor, such as by joining the nucleic acid coding for the polypeptide of interest with the nucleic acid sequence coding for the virulence factor and introducing the resulting gene construct into a cell capable of expressing 10 the conjugate.

Recombinant methodologies required to produce a DNA encoding 15 a desired protein are well known and routinely practiced in the art. Laboratory manuals, for example MOLECULAR CLONING: A LABORATORY MANUAL. Cold Spring Harbor Press: Cold Spring Harbor, N.Y. (1989) describes in detail techniques necessary to carry out the required DNA manipulations.

The fusion protein can be produced in recombinant microorganism transformed therewith. In this process, each protein component is preferably linked in the molecular ratio of 1:1 (injection domain : catalytic 20 domain). The aid of an appropriate linker, in order to allow proper folding of each protein molecule can be useful. As a linker, it is preferable to use a peptide consisting of the appropriate number of amino acids to maintain activity of each protein component, such as, a peptide composed of 0 to 20 amino acids, though glycine, (glycine)₄ serine, or 25 [(glycine)₄ serine]₂.

Preferable vectors include any of the well known prokaryotic expression vectors, recombinant baculoviruses, COS cell specific vectors, or yeast-specific expression constructs.

Alternatively, the two separate nucleotide sequences can be 30 expressed in a cell or can be synthesized chemically and subsequently joined, using known techniques. Alternatively, the fusion protein can be synthesized chemically as a single amino acid sequence (i.e., one in which both constituents are present) and, thus, joining is not needed.

35 Preferably, the injection domain of a Rho GTPase activator is a polypeptide selected from the group consisting of :

- a polypeptide comprising the amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 719 of sequence SEQ ID N°1;
- a polypeptide comprising the amino acid sequence starting at the 5 amino acid residue 1 and ending at the amino acid residue 719 of sequence SEQ ID N°2;
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 719 of sequence SEQ ID N°3; and
- 10 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 1145 of sequence SEQ ID N°4.

Preferably, the catalytic domain of a Rho GTPase activator is a polypeptide selected from the group consisting of :

- 15 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°1,
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°2,
- 20 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°3,
- a polypeptide comprising the amino acid sequence starting at the amino acid residue 1146 and ending at the amino acid residue 1451 of sequence SEQ ID N°4,
- 25 - a polypeptide comprising the amino acid sequence SEQ ID N°5,
- a polypeptide comprising the amino acid sequence SEQ ID N°6,
- a polypeptide comprising the amino acid sequence SEQ ID N°7,
- 30 - a polypeptide comprising the amino acid sequence SEQ ID N°8, and a polypeptide comprising the amino acid sequence SEQ ID N°9.

The invention concerns also the vaccine composition as described above, further comprising an antigen.

35 Preferably, the antigen is selected from the group consisting of a hormone, a protein, a drug, an enzyme, a vaccine composition against

bacterial, viral, fungal, prion, or parasitic infections, a component produced by microorganisms, inactivated bacterial toxins such as cholera toxin, ST and LT from *Escherichia coli*, tetanus toxin from *Clostridium tetani*, and proteins derived from HIV viruses.

5 The amount of antigen, and immunoadjuvant compound in the vaccine composition according to the invention, the dosages administered, are determined by techniques well known to those skilled in the pharmaceutical art, taking into consideration such factors as the particular antigen, the age, sex, weight, species, and condition of the
10 particular animal or patient, and the route of administration.

In a preferred embodiment, the vaccine composition according to the invention, further comprises one or more components selected from the group consisting of surfactants, absorption promoters, water absorbing polymers, substances which inhibit enzymatic degradation,
15 alcohols, organic solvents, oils, pH controlling agents, preservatives, osmotic pressure controlling agents, propellants, water and mixture thereof.

The vaccine composition according to the invention can further comprise a pharmaceutically acceptable carrier. The amount of the
20 carrier will depend upon the amounts selected for the other ingredients, the desired concentration of the antigen, the selection of the administration route, oral or parenteral, etc. The carrier can be added to the vaccine at any convenient time. In the case of a lyophilised vaccine, the carrier can, for example, be added immediately prior to
25 administration. Alternatively, the final product can be manufactured with the carrier.

Examples of appropriate carriers include, but are not limited to, sterile water, saline, buffers, phosphate-buffered saline, buffered sodium chloride, vegetable oils, Minimum Essential Medium (MEM), MEM with
30 HEPES buffer, etc.

Optionally, the vaccine composition of the invention may contain conventional, secondary adjuvants in varying amounts depending on the adjuvant and the desired result. The customary amount ranges from about 0.02% to about 20% by weight, depending upon the other ingredients and desired effect.
35

Examples of suitable secondary adjuvants include, but are not limited to, stabilizers; emulsifiers; aluminum hydroxide; aluminum phosphate; pH adjusters such as sodium hydroxide, hydrochloric acid, etc.; surfactants such as TweenTM 80 (polysorbate 80, commercially available from Sigma Chemical Co., St. Louis, Mo.); liposomes; iscom adjuvant; synthetic glycopeptides such as muramyl dipeptides; extenders such as dextran or dextran combinations, for example, with aluminum phosphate; carboxypolymethylene; bacterial cell walls such as mycobacterial cell wall extract; their derivatives such as Corynebacterium parvum; Propionibacterium acne; Mycobacterium bovis, for example, Bovine Calmette Guerin (BCG); vaccinia or animal poxvirus proteins; subviral particle adjuvants such as orbivirus; cholera toxin; N,N-di octadecyl-N',N'-bis(2-hydroxyethyl)-propanediamine (avridine); monophosphoryl lipid A; dimethyldioctadecylammonium bromide (DDA, commercially available from Kodak, Rochester, N.Y.); synthetics and mixtures thereof. Desirably, aluminum hydroxide is admixed with other secondary adjuvants or an immunoadjuvant such as Quil A.

Examples of suitable stabilizers include, but are not limited to, sucrose, gelatin, peptone, digested protein extracts such as NZ-Amine or NZ-Amine AS. Examples of emulsifiers include, but are not limited to, mineral oil, vegetable oil, peanut oil and other standard, metabolizable, nontoxic oils useful for injectables or intranasal vaccines compositions.

For the purpose of this invention, these adjuvants are identified herein as "secondary" merely to contrast with the above-described immunoadjuvant compound, consisting of a Rho GTPase activator, that is an essential ingredient in the vaccine composition for its effect in combination with an antigenic substance to significantly increase the humoral immune response to the antigenic substance. The secondary adjuvants are primarily included in the vaccine formulation as processing aids although certain adjuvants do possess immunologically enhancing properties to some extent and have a dual purpose.

Conventional preservatives can be added to the vaccine composition in effective amounts ranging from about 0.0001% to about 0.1% by weight. Depending on the preservative employed in the formulation, amounts below or above this range may be useful. Typical

preservatives include, for example, potassium sorbate, sodium metabisulfite, phenol, methyl paraben, propyl paraben, thimerosal, etc.

The choice of inactivated, modified or other type of vaccine composition and method of preparation of the improved vaccine composition formulation of the present invention are known or readily determined by those of ordinary skill in the art.

A pharmacologically effective amount of the immunoadjuvant compound according to the invention may be given, for example orally, parenterally or otherwise, concurrently with, sequentially to or shortly after the administration of a an antigenic substance in order to potentiate, accelerate or extend the immunogenicity of the antigen.

While the dosage of the vaccine composition depends upon the antigen, species, body weight of the host vaccinated or to be vaccinated, etc., the dosage of a pharmacologically effective amount of the vaccine composition will usually range from about 50 µg to about 500 µg per dose, per kilogram of body weight, in a mouse model.

Although the amount of the particular antigenic substance in the combination will influence the amount of the immunoadjuvant compound according to the invention, necessary to improve the immune response, it is contemplated that the practitioner can easily adjust the effective dosage amount of the immunoadjuvant compound through routine tests to meet the particular circumstances.

As a general rule, the vaccine composition of the present invention is conveniently administered orally, parenterally (subcutaneously, intramuscularly, intravenously, intradermally or intraperitoneally), intrabuccally, intranasally, or transdermally. The route of administration contemplated by the present invention will depend upon the antigenic substance and the co-formulants. For instance, if the vaccine composition contains saponins, while non-toxic orally or intranasally, care must be taken not to inject the sapogenin glycosides into the blood stream as they function as strong hemolytics. Also, many antigens will not be effective if taken orally. Preferably, the vaccine composition is administered subcutaneously, intramuscularly or intranasally.

The dosage of the vaccine composition will be dependent upon the selected antigen, the route of administration, species, body weight

and other standard factors. It is contemplated that a person of ordinary skill in the art can easily and readily titrate the appropriate dosage for an immunogenic response for each antigen to achieve the effective immunizing amount and method of administration.

5 The inventors have also shown, in example 1 that CNF1 has immunoadjuvant properties when coadministered orally with an antigen. They have also shown that this coadministration enhances the total IgA antibody titer in mice. This last result is typical of a mucosal response to an immunisation.

10 Consequently, a further object of the invention is a vaccine composition according to the invention, for administration to a mucosal surface.

15 This mode of administration presents a great interest. Indeed, the mucosal membranes contain numerous of dendritic cells and Langerhans cells, which are excellent antigen detecting and antigen presenting cells. The mucosal membranes are also connected to lymphoid organs called mucosal associated lymphoid tissue, which are able to forward an immune response to other mucosal areas. An example of such an epithelia is the nasal epithelial membrane, which consists of practically a 20 single layer of epithelial cells (pseudostratified epithelium) and the mucosal membrane in the upper respiratory tract is connected to the two lymphoid tissues, the adenoids and the tonsils. The extensive network of blood capillaries under the nasal mucosal of the high density of B and T cells, are particularly suited to provide a rapid recognition of the antigen 25 and provide a quick immunological response.

Preferably, the mucosal surface is selected from the group consisting of mucosal surfaces of the nose, lungs, mouth, eye, ear, gastrointestinal tract, genital tract, vagina, rectum, and the skin.

Another object of the invention is a vaccine composition for an oral 30 administration.

The invention concerns also a protein comprising a polypeptide consisting of; from the N-terminal end to the C-terminal end, respectively:

- a) the injection domain of a Rho GTPase activator as described above, and

- b) the catalytic domain of a Rho GTPase activator as described above.

The invention further concerns the use of a polypeptide of interest, for manufacturing a vaccine composition.

5 The invention also concerns the use of a fusion protein as described above for manufacturing a vaccine composition.

Further details of the invention are illustrated in the following non-limiting examples.

MATERIALS AND METHODS

10

Cells and reagents

Human umbilical vein endothelial cells (HUVEC) were obtained from PromoCell (Heidelberg, Germany). Cells were grown in Human Endothelial SFM medium (Invitrogen Co, Paisley, Scotland) 15 supplemented with defined growth factors (d-SFM): 10 ng/ml EGF and 20 ng/ml bFGF (Invitrogen Co), 1 µg/ml heparin (Sigma-Aldrich) and either 20% foetal bovine serum (Invitrogen Co) or 1% (W/V) bovine serum albumin (ELISA grade, Sigma-Aldrich) together with penicillin and streptomycin (Invitrogen Co). Cells were grown on 0,2% gelatine coated dishes (Sigma-Aldrich). Transfections of HUVEC were carried out as 20 described by Mettouchi et al., 2001. Antibodies used were monoclonal anti-β actin antibody [clone AC-74] (Sigma-Aldrich); anti-RhoA, anti-Cdc42, anti-Rac1 and anti-Ras antibodies (Transduction Laboratories); anti-HA [clone 11] (BabCO); anti-E-selectin [clone CTB202] (Santa Cruz Biotechnology) and rabbit polyclonal anti-phospho-p44/42 MAP kinase 25 (Thr202/Tyr204), anti phospho-p38 MAP kinase (Thr180/Tyr182) and anti phospho-c-Jun (Ser73) (Cell Signaling Technology); anti-human IκB-α (Upstate Biotechnology); anti-TRAF1 (H-186, Santa Cruz Biotechnology). Primary antibodies were visualized using goat anti- mouse or anti-rabbit horseradish peroxidase-conjugated secondary 30 antibodies (DAKO, Glostrup, Denmark). TRAF1 rabbit antibodies were visualized using biotin-XX goat anti-rabbit IgG followed by streptavidin horseradish peroxidase conjugate (Molecular Probes). DNA vectors corresponding to pcDNA3RhoQ63L, RacQ61L and Cdc42Q61L were 35 provided by Manor, D. (Lin et al., 1999).

Immunizations and immune response measurements.

Female BALB/c mice were purchased from Charles River Laboratories (L'Arbresle, France). Animals were maintained and handled according to
5 the regulations of the European Union and French Department of Health.
In all experiments 4-8 week-old female mice were immunized. Mice were fed with either CNF1 or CNF1-C866S or CT in the presence of 5 mg ovalbumin (OVA) (grade V, Sigma-Aldrich) dissolved in a solution of 500 μ l of 3% NaHCO₃. Levels of anti-OVA antibodies were determined by
10 means of solid-phase ELISA, as previously described by Anjuère et al., 2003. Mucosal anti-OVA IgA titers were determined according to the PERFEXT method (Villavedra et al., 1997). Briefly, guts were collected, weighted and homogenized 1:1 (W/V) in PBS supplemented with 2% saponin (SIGMA) and protease inhibitors (complete, Boehringer). IgA
15 titers were determined by means of solid-phase ELISA, as previously described (Anjuère et al., 2003). Antibodies used were goat anti-mouse IgG, IgG1, IgG2a, IgG2b and IgA, horseradish peroxidase conjugated (SouthernBiotech, Birmingham, USA).

20 DNA array analysis

HUVEC were seeded at 8 10⁶ cells/150 mm gelatin-coated dish in d-SFM containing BSA. Cells were intoxicated in parallel for 3h and 24h in d-SFM/BSA supplemented with 10⁻⁹M CNF1. Cells were lysed in RTL buffer for total RNA extraction, according to the manufacturer (RNeasy
25 MiniKit, Qiagen). CNF1 regulated genes were analyzed using Affymetrix® Human GeneChip U133A and U133B, by Aros Applied Biotechnology ApS (www.arosab.com), as recommended by the manufacturer (www.Affymetrix.com).

30 Toxins

Purified cholera toxin (CT) was obtained from List Biologicals (Campbell, CA). For CNF1 and CNF1-C866S toxin purification, *E. coli* OneShot, carrying pCR2cnf1 or pCR2cnf1C866S were grown overnight at 37°C in LB medium. Bacteria were harvested by centrifugation, suspended in 30 ml PBS and lysed using a French Press. The lysate was centrifuged 30

000 x g for 30 min, at 4°C and the supernatant was precipitated with the same volume of saturated ammonium sulphate for 5-8 hours. Precipitate was then dialysed against TN buffer (25 mM Tris [pH 7.4], 50 mM NaCl) and applied on a DEAE fast flow column (11 x 1.5 cm ; Pharmacia Biotech, Uppsala, Sweden). The column was washed for 200 min with the same buffer (1ml/min). CNF1 was eluted with a 50 to 300 mM NaCl gradient during 100 min (elution around 200 mM NaCl). The fractions containing CNF1 were pooled, dialysed against TN buffer and applied on a Superdex 75 column (0.3 ml/min ; Pharmacia Biotech). The fractions containing CNF1 were pooled, concentrated and applied on a monoQ column (Pharmacia Biotech). After a 20 min wash (1ml/min) of the column with TN buffer, CNF1 was eluted using a 50 to 400 mM NaCl gradient (elution around 350 mM NaCl). CNF1 purification was followed by SDS-PAGE. The activity of the different batches of CNF1 toxin was estimated by HEp-2 cells multinucleation assay, as previously described (Lemichez et al., 1997). The purified CNF1 toxin used in this study produced at 10^{-12} M 50% of multinucleation of HEp-2 cells after 48 hours of intoxication.

20 **ELISA**

HUVEC were seeded 24h before toxin addition at $2\ 10^5$ cells/ 22.5 mm or $5\ 10^5$ cells/ 35 mm well in d-SFM containing serum. Intoxication of cells was performed by addition of fresh medium containing CNF1, for different periods of time. One hour before intoxication ending the medium 25 was replaced by d-SFM containing BSA for ELISA. IL-8, MCP-1, IL-6, MIP3- α , TNF- α and RANTES production were assessed using human Quantikine® immunoassays, as recommended by the manufacturer (R & D Systems, Abingdon, UK).

30 **Pull-down and immunoblotting detection of activated-Rho GTPases**

Levels of activated-RhoA, -RhoB, -RhoC, -Rac1, -Rac2, -Cdc42 were measured using classical Rho effector pull-down assays developed by Manser et al., 1998 and Ren et al., 1999. For antibodies description 35 see the cells and reagents section.

Briefly, the measure of the levels of activated-RhoA, -B and -C was performed as followed. Cells were lysed in 50mM Tris, pH7.2, 500mM NaCl, 10mM MgCl₂, 1% Triton X-100, 0.5% deoxicholate, 0.1% SDS and protease inhibitors. Cell lysates were clarified by centrifugation
5 at 13000g at 4°C for 10min. and equal volumes of lysates (corresponding to 1mg of total proteins) were incubated with 30 micrograms GST-RBD (Rho binding domain of Rhotekin fused to GST and described in Ren et al., 1999) beads at 4°C for 45min. The beads were washed four times with buffer B (50mM Tris, pH7.2, 500mM NaCl, 10mM MgCl₂, 1% Triton
10 X-100 and protease inhibitors). Bound Rho proteins were resolved by SDS-PAGE and transferred on PVDF membranes. Activated-Rho proteins were detected by immunoblotting using a monoclonal antibody against either RhoA and RhoC or RhoB and anti-mouse horseradish peroxidase-conjugated secondary antibody followed by
15 chemiluminescence detection.

The measure of the levels of activated-Rac1, Rac2 and Cdc42 was determined, as followed. Cells were lysed in LB buffer (25 mM Tris, pH7.5, 150mM NaCl, 5mM MgCl₂, 0.5% Triton X-100, 4% glycerol and protease inhibitors). Cell lysates were clarified by centrifugation at
20 13000g at 4°C for 10min. and equal volumes of lysates (corresponding to 1mg of total proteins) were incubated with 30 micrograms GST-PAK70-106 (Rac/Cdc42 binding domain of p21PAK fused to GST and described in Manser et al., 1998) beads at 4°C for 45min. The beads were washed four times with LB. Bound Rac and Cdc42 proteins were resolved by
25 SDS-PAGE and transferred on PVDF membranes. Activated-Rac1, 2 or activated-Cdc42 proteins were detected by immunoblotting using a monoclonal antibody against either Rac1, 2 or Cdc42 and anti-mouse horseradish peroxidase-conjugated secondary antibody followed by chemiluminescence detection.

30 For activated Ras measurements GST-RBD1-149 of Raf1 was used as described by the authors (de Rooij and Bos, 1997).

Example 1: CNF1 effects on cell signaling pathways

35 Kinetics of CNF1-induced Rac1, Cdc42 and RhoA activation have been

studied. These kinetics show the specificity of Rho protein activation, as compared to the Ras GTPase (Fig. 1A, 1B). Obviously, these measurements do not represent an exhaustive list of the Rho proteins activated by CNF1, other Rho bearing the canonical sequence for CNF1
5 recognition/modification (Lerm et al., 1999). These measurements rather indicated that all the three Rho proteins exhibited a maximal activation around 2 hours in HUVEC intoxicated with 10^{-9} M CNF1 (Fig. 1B). CNF1 interference with classical signaling pathways leading to gene regulation, has also been shown. Consistent with the absence of Ras activation
10 measured, CNF1 did not produce ERK1/2 phosphorylation (Fig. 1A, 1C). CNF1 rather appeared to interfere both with the SAP-kinase signaling pathways, unraveled by p38MAP-kinase and cjun phosphorylations. CNF1 also interferes with the NF-kappaB pathway, as shown by I κ B depletion (Fig. 1C). Host cells have evolved cell surface receptors to get
15 alarmed of the presence of PAMP (Medzhitov and Janeway, 2002). PAMP receptors initiate an innate immune response through I κ B depletion for NF κ B activation (Barton and Medzhitov, 2003). That cell treatment with the catalytic inactive CNF1-C866S toxin was devoid of interference with all signaling pathways tested, especially NF κ B, strongly
20 suggested an absence of cell recognition of CNF1 as a PAMP (Fig. 1C).

Example 2 : CNF1 potentiates host adaptative responses to a bystander antigen.

25 The systemic and mucosal humoral immune responses in mice fed CNF1 together with ovalbumin (OVA), a prototype soluble protein antigen, has been studied. Oral co-administration of OVA and CNF1 induced serum IgG antibody responses to OVA that were almost comparable in magnitude to those evoked by cholera toxin (CT) (Fig.
30 2A), the most potent mucosal adjuvant (Holmgren et al., 2003). These responses were comprised predominantly of IgG1 and IgG2b, indicating a bias toward a classical Th-2 response (Fig 2B). Feeding OVA alone failed to induce detectable antibody responses (not shown). Furthermore, feeding mice CNF1 evoked strong IgA antibody responses to co-administered OVA in the small intestinal mucosal (Fig. 2C). Taken
35

together, these results demonstrate that CNF1 displays adjuvant properties on gut-induced mucosal and systemic immune responses. To examine whether CNF1 catalytic activity was required to support the adjuvanticity of this toxin, the effects of wild type CNF1 to that of the enzymatically inactive mutant (CNF1-C866S) have been compared. The results show that the enzymatic activity of CNF1 is necessary to promote its adjuvanticity for systemic as well as mucosal anti-OVA antibody responses (Fig. 2). In conclusion, this study indicates that CNF1-induced activation of Rho proteins efficiently triggers a host cell alarm program and suggests that this toxin is endowed immunomodulatory properties on innate and adaptative immune responses.

Example 3 The catalytic domain of DNT remains active on cells and is sufficient to confer adjuvanticity

CNF1 belongs to a family of toxins among them DNT, having similar catalytic activity (Boquet and Lemichez 2003). It is shown on Figure 3A that the catalytic domain of DNT (DNT-CTER) remains active on cells, although showing a lower intoxication property as compared to CNF1. Despite its inability to intoxicate cells (Fig. 3A), the catalytic domain of CNF1 (CNF1-CTER) upon mechanical injection into cells produces a *bona fide* toxic phenotype (Lemichez et al., 1997). It has been taken advantage of the above observations to test the adjuvant properties of the catalytic domains of both toxins. Mice were fed 10 times higher quantities of both toxin catalytic domains, as compared to CNF1. In these conditions it has been observed that DNT-CTER stimulated significantly the anti-OVA IgG responses (Fig. 3B). CNF1-CTER also produced a stimulation of the anti-OVA IgG responses, although at a lower level (Fig. 3B). Taken together, these results indicate that the adjuvanticity of this group of toxin is encompassed in their catalytic domain. Nevertheless, the injection domain of CNF1-toxin together with its catalytic domain, allows the use of lower doses to induce a significantly higher biological effect.

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Claims

1. A vaccine composition comprising an immunoadjuvant compound, wherein said immunoadjuvant compound consists of a Rho GTPase activator.
5
2. A vaccine composition according to claim 1 wherein said immunoadjuvant compound is selected from the group consisting of :
 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°1,
 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°2,
 - 10 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°3,
 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 1146 and ending at the amino acid residue 1451
15 of sequence SEQ ID N°4,
 - a polypeptide comprising the amino acid sequence SEQ ID N°5,
 - a polypeptide comprising the amino acid sequence SEQ ID N°6,
 - a polypeptide comprising the amino acid sequence SEQ ID N°7,
 - a polypeptide comprising the amino acid sequence SEQ ID N°8, and
20
 - a polypeptide comprising the amino acid sequence SEQ ID N°9.
3. A vaccine composition according to claim 1 wherein said immunoadjuvant compound is selected from the group consisting of :
30
 - a polypeptide comprising the amino acid sequence SEQ ID N°1,
 - a polypeptide comprising the amino acid sequence SEQ ID N°2,
 - a polypeptide comprising the amino acid sequence SEQ ID N°3, and
 - a polypeptide comprising the amino acid sequence SEQ ID N°4.

4. A vaccine composition according to claim 1, wherein said immunoadjuvant compound is a protein comprising a polypeptide consisting of; from the N-terminal end to the C-terminal end, respectively:
 - 5 a) the injection domain of a Rho GTPase activator , and
 - b) the catalytic domain of a Rho GTPase activator.
- 10 5. A vaccine composition according to claim 4, wherein said injection domain of a Rho GTPase activator is a polypeptide selected from the group consisting of :
 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 719 of sequence SEQ ID N°1;
 - a polypeptide comprising the amino acid sequence starting at the 15 amino acid residue 1 and ending at the amino acid residue 719 of sequence SEQ ID N°2;
 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 719 of sequence SEQ ID N°3; and
 - 20 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 1 and ending at the amino acid residue 1145 of sequence SEQ ID N°4.
- 25 6. A vaccine composition according to anyone of claims 4 and 5, wherein said catalytic domain of a Rho GTPase activator is a polypeptide selected from the group consisting of :
 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°1,
 - 30 - a polypeptide comprising the amino acid sequence starting at the amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°2,
 - a polypeptide comprising the amino acid sequence starting at the 35 amino acid residue 720 and ending at the amino acid residue 1014 of sequence SEQ ID N°3,

- a polypeptide comprising the amino acid sequence starting at the amino acid residue 1146 and ending at the amino acid residue 1451 of sequence SEQ ID N°4,
 - a polypeptide comprising the amino acid sequence SEQ ID N°5,
 - 5 - a polypeptide comprising the amino acid sequence SEQ ID N°6,
 - a polypeptide comprising the amino acid sequence SEQ ID N°7,
 - a polypeptide comprising the amino acid sequence SEQ ID N°8, and
 - a polypeptide comprising the amino acid sequence SEQ ID N°9.
- 10 7. A vaccine composition according to anyone of claim 1-6 comprising further an antigen.
8. A vaccine composition according to claim 7 wherein the antigen is selected from the group consisting of a hormone, a protein, a drug, an enzyme, a vaccine composition against bacterial, viral, fungal, prion, or parasitic infections, a component produced by microorganisms, inactivated bacterial toxins such as cholera toxin, ST and LT from *Escherichia coli*, tetanus toxin from *Clostridium tetani*, and proteins derived from HIV viruses.
- 15 20 9. A vaccine composition according to anyone of claims 1-8 for administration to a mucosal surface.
- 25 10. A vaccine composition according to anyone of claims 1-9, for an oral administration.
11. A protein comprising a polypeptide consisting of; from the N-terminal end to the C-terminal end, respectively :
30 a) the injection domain of a Rho GTPase activator according to anyone of claims 4 and 5, and
b) the catalytic domain of a Rho GTPase activator according to anyone of claims 4 and 6.
- 35 12. Use of a polypeptide as defined in anyone of claim 1-6 for manufacturing a vaccine composition.

13. Use of a protein according to claim 11 for manufacturing a vaccine composition.

A B S T R A C T O F D I S C L O S U R E

A vaccine composition comprising an Immunoadjuvant compound consisting of a Rho GTPase family activator

INSTITUT NATIONAL DE LA SANTE ET DE LA
RECHERCHE MEDICALE INSERM

This invention is based on the experimental finding that activators of Rho GTPases, namely the cytotoxic necrotizing factor 1 (CNF1), and DNT bear immunostimulatory properties towards the systemic response to orally administered ovalbumine.

This invention concerns a vaccine composition comprising an immunoadjuvant compound, wherein said immunoadjuvant compound consists of a Rho GTPase activator.



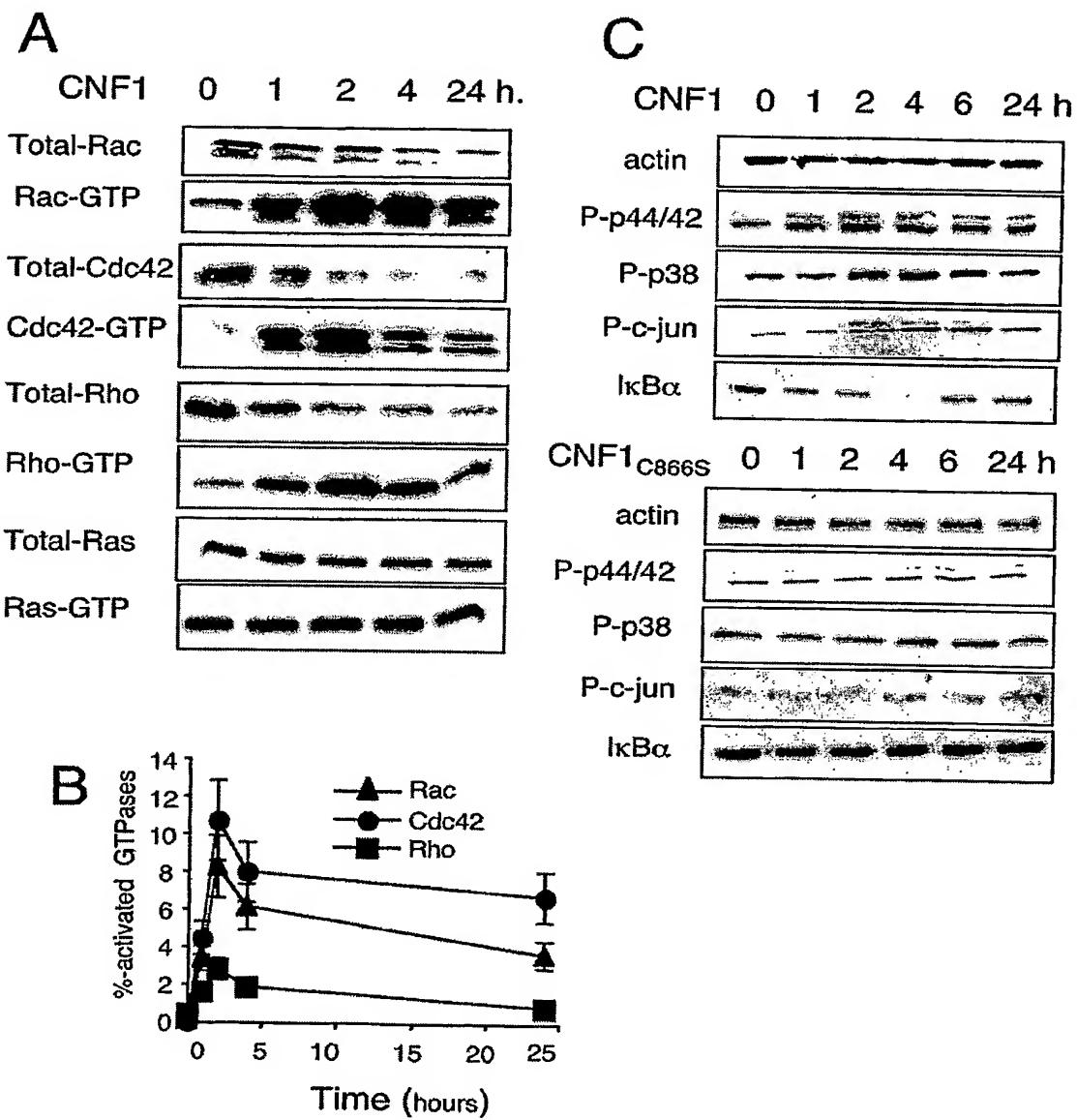


FIGURE 1

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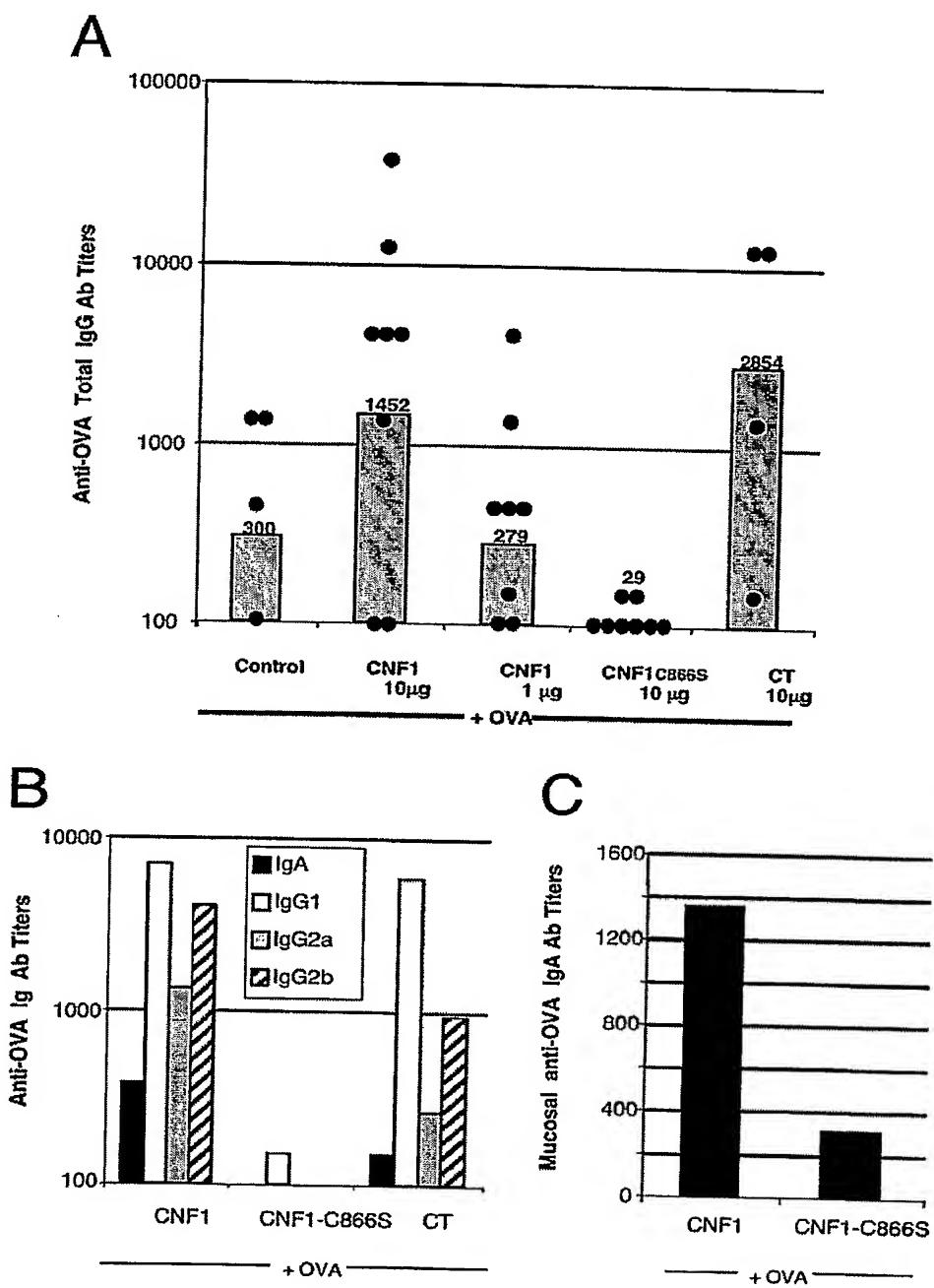
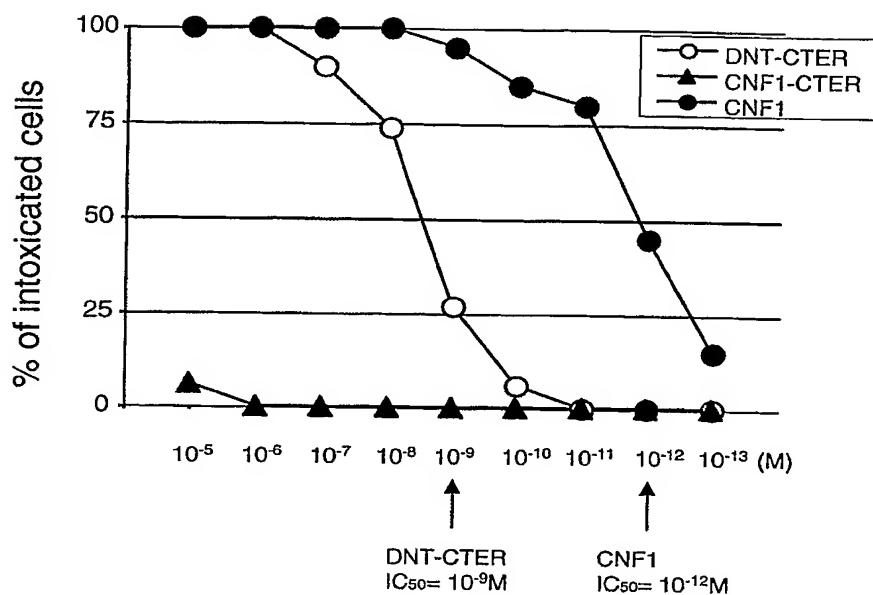


FIGURE 2

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A



B

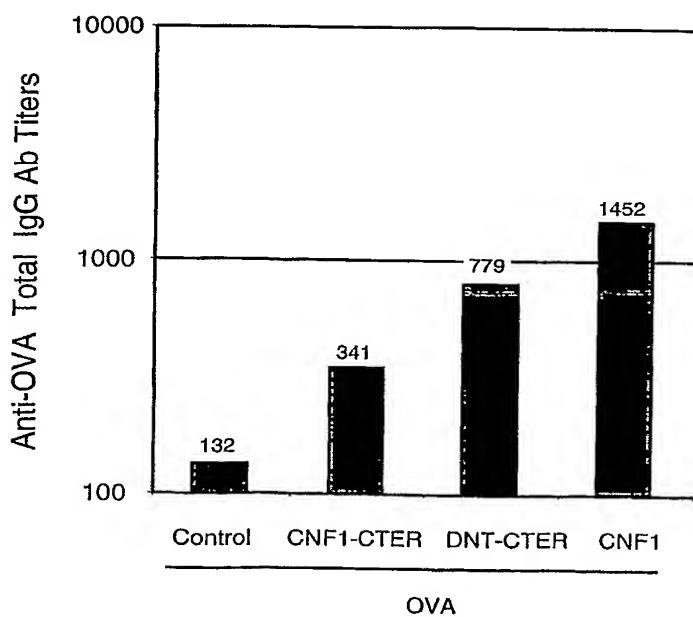
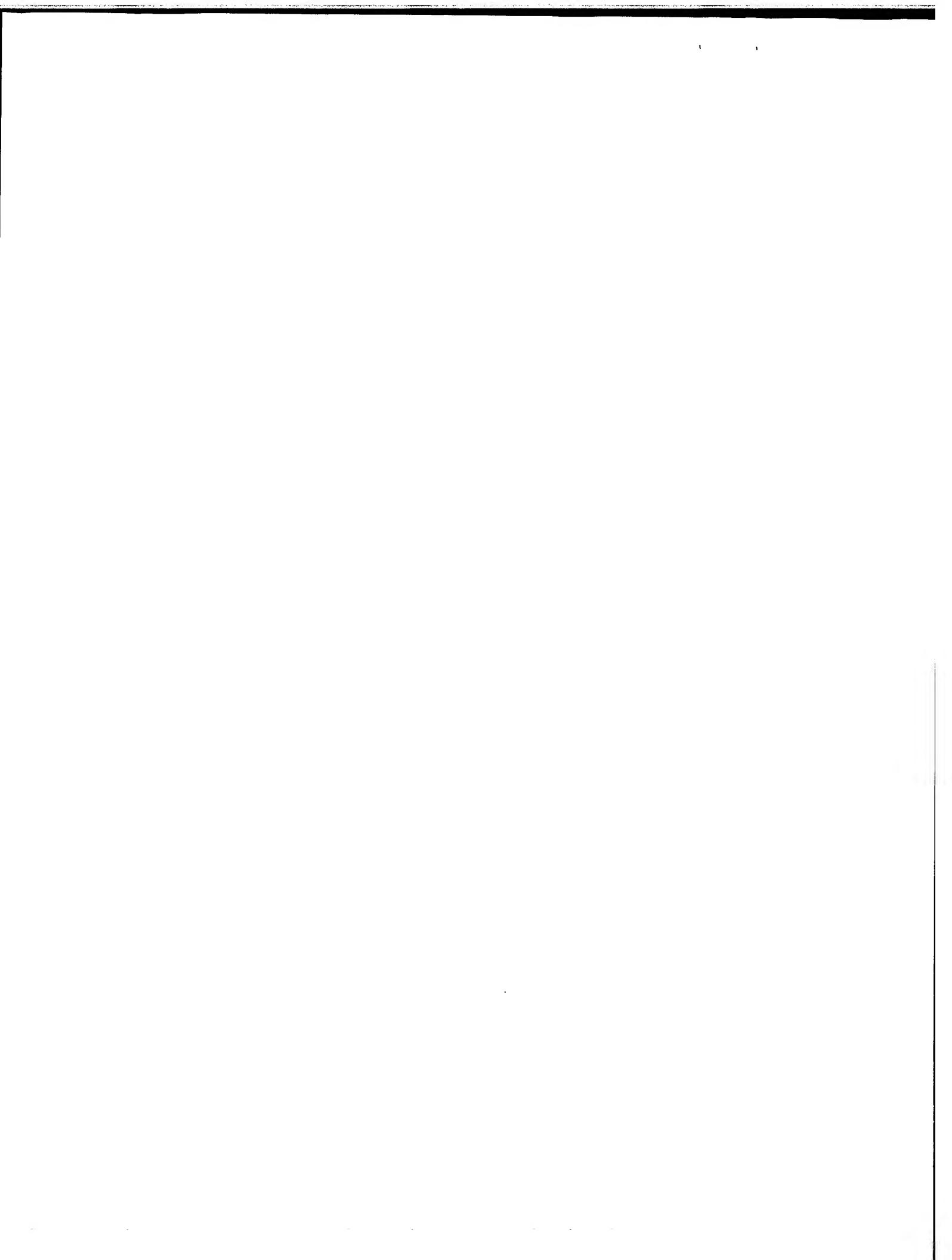


FIGURE 3



SEQUENCE LISTING

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<170> PatentIn version 3.1

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Leu Thr Gln Ala Ala Leu Ile Asn Ile Gly Tyr Arg Phe Asp Val Phe
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Asp Asp Ala Asn Ser Ser Thr Gly Ile Tyr Lys Thr Lys Ser Ala Asp
100 105 110

Val Phe Asn Glu Glu Asn Glu Glu Lys Met Leu Pro Ser Glu Tyr Leu
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His Phe Leu Gln Lys Cys Asp Phe Ala Gly Val Tyr Gly Lys Thr Leu
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Ser Asp Tyr Trp Ser Lys Tyr Tyr Asp Lys Phe Lys Leu Leu Leu Lys
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Asn Tyr Tyr Ile Ser Ser Ala Leu Tyr Leu Tyr Lys Asn Gly Glu Leu

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Asp Glu Arg Glu Tyr Asn Phe Ser Met Asn Ala Leu Asn Arg Ser Asp
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Asn Ile Ser Leu Leu Phe Phe Asp Ile Tyr Gly Tyr Tyr Ala Ser Asp
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Ile Phe Val Ala Lys Asn Asn Asp Lys Val Met Leu Phe Ile Pro Gly
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Ala Lys Lys Pro Phe Leu Phe Lys Lys Asn Ile Ala Asp Leu Arg Leu
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Thr Leu Lys Glu Leu Ile Lys Asp Ser Asp Lys Gln Gln Leu Leu Ser
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Gln His Phe Ser Leu Tyr Ser Arg Gln Asp Gly Val Ser Tyr Ala Gly
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Val Asn Ser Val Leu His Ala Ile Glu Asn Asp Gly Asn Phe Asn Glu
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Ser Tyr Phe Leu Tyr Ser Asn Lys Thr Leu Ser Asn Lys Asp Val Phe
290 295 300

Asp Ala Ile Ala Ile Ser Val Lys Lys Arg Ser Phe Ser Asp Gly Asp
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Gln Tyr Phe Tyr Asp Asn Thr Val Gly Leu Asn Gly Ile Pro Thr Leu
785 790 795 800

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Pro Arg Val Glu Gly Ile Met Ser Asn Asp Phe Leu Val Asp Tyr Leu
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Ser Glu Asn Phe Glu Asp Ser Leu Ile Thr Tyr Ser Ser Ser Glu Lys
930 935 940

Lys Pro Asp Ser Gln Ile Thr Ile Arg Asp Asn Val Ser Val Phe
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Pro Tyr Phe Leu Asp Asn Ile Pro Glu His Gly Phe Gly Thr Ser Ala
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Thr Val Leu Val Arg Val Asp Gly Asn Val Val Val Arg Ser Leu Ser
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Asp Asp Ala Asn Ala Ser Ala Gly Ile Tyr Lys Thr Ser Ser Ala Asp
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Met Phe Asn Glu Lys Asn Glu Glu Lys Met Leu Pro Ser Glu Tyr Leu
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Asp Glu Tyr Glu Tyr Asn Phe Ser Ile Ser Ala Leu Asn Arg Arg Asp
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Asn Ile Ser Leu Phe Phe Asp Ile Tyr Gly Tyr Tyr Ser Ser Asp
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Asp Glu Ala Asn Ser Thr Ala Gly Ile Tyr Lys Thr Asn Asn Ala Asp
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Ser Phe Asp Glu Thr Asn Glu Ala Lys Met Leu Pro Ser Glu Tyr Leu
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Pro Leu Leu Phe Pro Glu Met Asp Ser Arg Leu Pro Lys Pro Thr Pro
565 570 575

Glu Leu Asp Ile Lys Tyr Tyr Ser Ser Asn Leu Ser Ser Phe Lys Glu
580 585 590

Asp Thr Val Ile Leu Met Arg Gly Thr Thr Glu Glu Ala Trp Asn
595 600 605

Ile Ala Asn Tyr Lys Thr Ala Gly Gly Ser Asn Lys Asp Leu Glu Glu
610 615 620

Asn Phe Ile Glu Ala Gly Pro Gln Phe Asn Leu Ser Phe Ser Glu Tyr
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Thr Ser Ser Ile Asn Ser Ala Asp Thr Ala Ser Arg Lys His Phe Leu
645 650 655

Val Ile Ile Lys Val Gln Val Lys Tyr Ile Ser Asn Asp Asn Val Leu
660 665 670

Tyr Ala Asn His Trp Ala Ile Pro Asp Glu Ala Pro Val Glu Val Leu
675 680 685

Ala Val Val Asp Arg Arg Phe Ile Phe Pro Glu Pro Pro Val Lys Pro
690 695 700

Lys Leu Ser Phe Ile Gln Lys Ile Ala Asn Arg Phe Leu Thr Glu Asn
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740 745 750

Glu Ile Tyr Leu Arg Phe Asp Ala Ala Asn Ala Asp Glu Leu Arg Pro
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Gly Asp Val Tyr Val Lys Lys Thr Lys Phe Asp Ser Met Gly Tyr Asp
770 775 780

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785 790 795 800

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Thr Tyr Trp Leu Lys Tyr Asn Leu Thr Asn Glu Thr Ser Ile Ile Lys
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Glu Ile Glu Glu Asn Lys Pro Val Val Ile Thr Ser Gly Thr Leu Thr
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Gly Cys Thr Val Val Phe Ala Arg Lys Gly Glu Tyr Phe Tyr Ala Val
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His Thr Gly Asn Ser Glu Ser Leu Ile Gly Phe Thr Ser Thr Ser Gly
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915 920 925

Ser Asp Asn Phe Asp Ser Ala Leu Ile Ser Tyr Ser Ser Ser Ser Leu
930 935 940

Lys Pro Asn Ser Met Ile Asn Ile Ser Arg Glu Asn Val Ser Thr Phe
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Ser Tyr Tyr Thr Asp Asp Ile Gln Leu Pro Ser Phe Gly Thr Ser Val
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Leu Val Phe Glu Tyr Val Lys Ala Arg Tyr Glu Ile Tyr Tyr Leu Leu
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Gly Val Leu Ala Arg Phe Arg Arg Leu Glu Gln Glu Thr Ala Gly Met
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Asp Phe Arg Asp Ser Asp Ala Phe Ala Ser Tyr Ala Glu Tyr Ala Ala
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Gln Phe Asn Asp Tyr Ile Asp Gln Tyr Ser Ile Leu Glu Ala Gln Arg
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995 1000 1005

Tyr Asp Leu Asp Asp Val Ala Thr Leu Phe Asn Ala Val Asp Arg
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Asn Thr Ser Leu Gly Arg Gln Ala Arg Met Glu Leu Tyr Leu Asp
1025 1030 1035

Ala Ile Val Asp Leu His Ala Arg Leu Gly Tyr Glu Asn Ala Arg
1040 1045 1050

Phe Val Asp Leu Met Ala Phe His Leu Leu Ser Leu Gly His Ala
1055 1060 1065

Ala Thr Ala Ser Glu Val Val Glu Ala Val Ser Pro Arg Leu Leu
1070 1075 1080

Gly Asn Val Phe Asp Ile Ser Asn Val Ala Gln Leu Glu Arg Gly
1085 1090 1095

Ile Gly Asn Pro Ala Ser Thr Gly Leu Phe Val Met Leu Gly Ala
1100 1105 1110

Tyr Ser Glu Ser Ser Pro Ala Ile Phe Gln Ser Phe Val Asn Asp
1115 1120 1125

Ile Phe Pro Ala Trp Arg Gln Ala Ser Gly Gly Pro Leu Val
1130 1135 1140

Trp Asn Phe Gly Pro Ala Ala Ile Ser Pro Thr Arg Leu Asp Tyr

1145

1150

1155

Ala Asn Thr Asp Ile Gly Leu Leu Asn His Gly Asp Ile Ser Pro
1160 1165 1170

Leu Arg Ala Arg Pro Pro Leu Gly Gly Arg Arg Asp Ile Asp Leu
1175 1180 1185

Pro Pro Gly Leu Asp Ile Ser Phe Val Arg Tyr Asp Arg Pro Val
1190 1195 1200

Arg Met Ser Ala Pro Arg Ala Leu Asp Ala Ser Val Phe Arg Pro
1205 1210 1215

Val Asp Gly Pro Val His Gly Tyr Ile Gln Ser Trp Thr Gly Ala
1220 1225 1230

Glu Ile Glu Tyr Ala Tyr Gly Ala Pro Ala Ala Ala Arg Glu Val
1235 1240 1245

Met Leu Thr Asp Asn Val Arg Ile Ile Ser Ile Glu Asn Gly Asp
1250 1255 1260

Glu Gly Ala Ile Gly Val Arg Val Arg Leu Asp Thr Val Pro Val
1265 1270 1275

Ala Thr Pro Leu Ile Leu Thr Gly Gly Ser Leu Ser Gly Cys Thr
1280 1285 1290

Thr Met Val Gly Val Lys Glu Gly Tyr Leu Ala Phe Tyr His Thr
1295 1300 1305

Gly Lys Ser Thr Glu Leu Gly Asp Trp Ala Thr Ala Arg Glu Gly
1310 1315 1320

Val Gln Ala Leu Tyr Gln Ala His Leu Ala Met Gly Tyr Ala Pro
1325 1330 1335

Ile Ser Ile Pro Ala Pro Met Arg Asn Asp Asp Leu Val Ser Ile
1340 1345 1350

Ala Ala Thr Tyr Asp Arg Ala Val Ile Ala Tyr Leu Gly Lys Asp
1355 1360 1365

Val Pro Gly Gly Ser Thr Arg Ile Thr Arg His Asp Glu Gly
1370 1375 1380

Ala Gly Ser Val Val Ser Phe Asp Tyr Asn Ala Ala Val Gln Ala
1385 1390 1395

Ser Ala Val Pro Arg Leu Gly Gln Val Tyr Val Leu Ile Ser Asn
1400 1405 1410

Asp Gly Gln Gly Ala Arg Ala Val Leu Leu Ala Glu Asp Leu Ala
1415 1420 1425

Trp Ala Gly Ser Gly Ser Ala Leu Asp Val Leu Asn Glu Arg Leu
1430 1435 1440

Val Thr Leu Phe Pro Ala Pro Val
1445 1450

<210> 5
<211> 240
<212> PRT
<213> *Salmonella typhimurium*

<400> 5

Met Thr Lys Ile Thr Leu Ser Pro Gln Asn Phe Arg Ile Gln Lys Gln
1 5 10 15

Glu Thr Thr Leu Leu Lys Glu Lys Ser Thr Glu Lys Asn Ser Leu Ala
20 25 30

Lys Ser Ile Leu Ala Val Lys Asn His Phe Ile Glu Leu Arg Ser Lys
35 40 45

Leu Ser Glu Arg Phe Ile Ser His Lys Asn Thr Glu Ser Ser Ala Thr
50 55 60

His Phe His Arg Gly Ser Ala Ser Glu Gly Arg Ala Val Leu Thr Asn
65 70 75 80

Lys Val Val Lys Asp Phe Met Leu Gln Thr Leu Asn Asp Ile Asp Ile
85 90 95

Arg Gly Ser Ala Ser Lys Asp Pro Ala Tyr Ala Ser Gln Thr Arg Glu
100 105 110

Ala Ile Leu Ser Ala Val Tyr Ser Lys Asn Lys Asp Gln Cys Cys Asn
115 120 125

Leu Leu Ile Ser Lys Gly Ile Asn Ile Ala Pro Phe Leu Gln Glu Ile
130 135 140

Gly Glu Ala Ala Lys Asn Ala Gly Leu Pro Gly Thr Thr Lys Asn Asp
145 150 155 160

Val Phe Thr Pro Ser Gly Ala Gly Ala Asn Pro Phe Ile Thr Pro Leu
165 170 175

Ile Ser Ser Ala Asn Ser Lys Tyr Pro Arg Met Phe Ile Asn Gln His
180 185 190

Gln Gln Ala Ser Phe Lys Ile Tyr Ala Glu Lys Ile Ile Met Thr Glu
195 200 205

Val Ala Pro Leu Phe Asn Glu Cys Ala Met Pro Thr Pro Gln Gln Phe
210 215 220

Gln Leu Ile Leu Glu Asn Ile Ala Asn Lys Tyr Ile Gln Tyr Thr Pro
225 230 235 240

<210> 6
<211> 240
<212> PRT
<213> *Salmonella typhimurium*

<400> 6

Met Thr Asn Ile Thr Leu Ser Thr Gln His Tyr Arg Ile His Arg Ser
1 5 10 15

Asp Val Glu Pro Val Lys Glu Lys Thr Thr Glu Lys Asp Ile Phe Ala
20 25 30

Lys Ser Ile Thr Ala Val Arg Asn Ser Phe Ile Ser Leu Ser Thr Ser
35 40 45

Leu Ser Asp Arg Phe Ser Leu His Gln Gln Thr Asp Ile Pro Thr Thr
50 55 60

His Phe His Arg Gly Asn Ala Ser Glu Gly Arg Ala Val Leu Thr Ser
65 70 75 80

Lys Thr Val Lys Asp Phe Met Leu Gln Lys Leu Asn Ser Leu Asp Ile
85 90 95

Lys Gly Asn Ala Ser Lys Asp Pro Ala Tyr Ala Arg Gln Thr Cys Glu
100 105 110

Ala Ile Leu Ser Ala Val Tyr Ser Asn Asn Lys Asp Gln Cys Cys Lys
115 120 125

Leu Leu Ile Ser Lys Gly Val Ser Ile Thr Pro Phe Leu Lys Glu Ile
130 135 140

Gly Glu Ala Ala Gln Asn Ala Gly Leu Pro Gly Glu Ile Lys Asn Gly
145 150 155 160

Val Phe Thr Pro Gly Gly Ala Gly Ala Asn Pro Phe Val Val Pro Leu
165 170 175

Ile Ala Ser Ala Ser Ile Lys Tyr Pro His Met Phe Ile Asn His Asn
180 185 190

Gln Gln Val Ser Phe Lys Ala Tyr Ala Glu Lys Ile Val Met Lys Glu
195 200 205

Val Thr Pro Leu Phe Asn Lys Gly Thr Met Pro Thr Pro Gln Gln Phe
210 215 220

Gln Leu Thr Ile Glu Asn Ile Ala Asn Lys Tyr Leu Gln Asn Ala Ser
225 230 235 240

<210> 7
<211> 166
<212> PRT
<213> Shigella flexneri

<400> 7

Met Glu Ile Gln Asn Thr Lys Ser Ala Pro Ile Leu Tyr Thr Asp Ile
1 5 10 15

Ser Thr Lys Gln Thr Gln Ser Ser Glu Thr Gln Lys Ser Gln Asn
20 25 30

Tyr Gln Gln Leu Ala Ala His Ile Pro Leu Asn Val Gly Lys Asn Pro
35 40 45

Val Leu Thr Thr Thr Leu Asn Asp Asp Gln Leu Leu Lys Leu Ser Glu
50 55 60

Gln Val Gln His Asp Ser Glu Ile Ile Ala Arg Leu Thr Asp Lys Lys
65 70 75 80

Met Lys Asp Leu Ser Glu Met Ser His Thr Ile Thr Pro Glu Asn Thr
85 90 95

Leu Asp Ile Ser Ser Leu Ser Ser Asn Ala Val Ser Leu Ile Ile Ser
100 105 110

Val Ala Val Leu Leu Ser Ala Leu Arg Thr Ala Glu Thr Arg Leu Gly
115 120 125

Ser Gln Leu Ser Leu Ile Ala Phe Asp Ala Thr Lys Ser Ala Ala Glu
130 135 140

Asn Ile Val Arg Gln Gly Leu Ala Ala Leu Ser Ser Ser Ile Thr Gly
145 150 155 160

Ala Val Thr Gln Val Gly
165

<210> 8
<211> 1186
<212> PRT
<213> Helicobacter pylori

<400> 8

Met Thr Asn Glu Thr Ile Asp Gln Thr Arg Thr Pro Asp Gln Thr Gln
1 5 10 15

Ser Gln Thr Ala Phe Asp Pro Gln Phe Ile Asn Asn Leu Gln Val
20 25 30

Ala Phe Ile Lys Val Asp Asn Val Val Ala Ser Phe Asp Pro Asp Gln
35 40 45

Lys Pro Ile Val Asp Lys Asn Asp Arg Asp Asn Arg Gln Ala Phe Asp
50 55 60

Gly Ile Ser Gln Leu Arg Glu Glu Tyr Ser Asn Lys Ala Ile Lys Asn
65 70 75 80

Pro Thr Lys Lys Asn Gln Tyr Phe Ser Asp Phe Ile Asp Lys Ser Asn
85 90 95

Asp Leu Ile Asn Lys Asp Asn Leu Ile Asp Val Glu Ser Ser Thr Lys
100 105 110

Ser Phe Gln Lys Phe Gly Asp Gln Arg Tyr Gln Ile Phe Thr Ser Trp
115 120 125

Val Ser His Gln Lys Asp Pro Ser Lys Ile Asn Thr Arg Ser Ile Arg
130 135 140

Asn Phe Met Glu Asn Ile Ile Gln Pro Pro Ile Pro Asp Asp Lys Glu
145 150 155 160

Lys Ala Glu Phe Leu Lys Ser Ala Lys Gln Ser Phe Ala Gly Ile Ile
165 170 175

Ile Gly Asn Gln Ile Arg Thr Asp Gln Lys Phe Met Gly Val Phe Asp
180 185 190

Glu Ser Leu Lys Glu Arg Gln Glu Ala Glu Lys Asn Gly Gly Pro Thr
195 200 205

Gly Gly Asp Trp Leu Asp Ile Phe Leu Ser Phe Ile Phe Asn Lys Lys
210 215 220

Gln Ser Ser Asp Val Lys Glu Ala Ile Asn Gln Glu Pro Val Pro His
225 230 235 240

Val Gln Pro Asp Ile Ala Thr Thr Thr Asp Ile Gln Gly Leu Pro
245 250 255

Pro Glu Ala Arg Asp Leu Leu Asp Glu Arg Gly Asn Phe Ser Lys Phe
260 265 270

Thr Leu Gly Asp Met Glu Met Leu Asp Val Glu Gly Val Ala Asp Ile
275 280 285

Asp Pro Asn Tyr Lys Phe Asn Gln Leu Leu Ile His Asn Asn Ala Leu
290 295 300

Ser Ser Val Leu Met Gly Ser His Asn Gly Ile Glu Pro Glu Lys Val
305 310 315 320

Ser Leu Leu Tyr Ala Gly Asn Gly Gly Phe Gly Asp Lys His Asp Trp
325 330 335

Asn Ala Thr Val Gly Tyr Lys Asp Gln Gln Gly Asn Asn Val Ala Thr
340 345 350

Leu Ile Asn Val His Met Lys Asn Gly Ser Gly Leu Val Ile Ala Gly
355 360 365

Gly Glu Lys Gly Ile Asn Asn Pro Ser Phe Tyr Leu Tyr Lys Glu Asp
370 375 380

Gln Leu Thr Gly Ser Gln Arg Ala Leu Ser Gln Glu Glu Ile Arg Asn
385 390 395 400

Lys Val Asp Phe Met Glu Phe Leu Ala Gln Asn Asn Thr Lys Leu Asp
405 410 415

Asn Leu Ser Glu Lys Glu Lys Phe Gln Asn Glu Ile Glu Asp
420 425 430

Phe Gln Lys Asp Ser Lys Ala Tyr Leu Asp Ala Leu Gly Asn Asp Arg
435 440 445

Ile Ala Phe Val Ser Lys Lys Asp Thr Lys His Ser Ala Leu Ile Thr
450 455 460

Glu Phe Asn Asn Gly Asp Leu Ser Tyr Thr Leu Lys Asp Tyr Gly Lys
465 470 475 480

Lys Ala Asp Lys Ala Leu Asp Arg Glu Lys Asn Val Thr Leu Gln Gly
485 490 495

Ser Leu Lys His Asp Gly Val Met Phe Val Asp Tyr Ser Asn Phe Lys
500 505 510

Tyr Thr Asn Ala Ser Lys Asn Pro Asn Lys Gly Val Gly Ala Thr Asn
515 520 525

Gly Val Ser His Leu Glu Ala Gly Phe Asn Lys Val Ala Val Phe Asn
530 535 540

Leu Pro Asp Leu Asn Asn Leu Ala Ile Thr Ser Phe Val Arg Arg Asn
545 550 555 560

Leu Glu Asn Lys Leu Thr Ala Lys Gly Leu Ser Leu Gln Glu Ala Asn
565 570 575

Lys Leu Ile Lys Asp Phe Leu Ser Ser Asn Lys Glu Leu Ala Gly Lys
580 585 590

Ala Leu Asn Phe Asn Lys Ala Val Ala Glu Ala Lys Ser Thr Gly Asn
595 600 605

Tyr Asp Glu Val Lys Lys Ala Gln Lys Asp Leu Glu Lys Ser Leu Arg
610 615 620

Lys Arg Glu His Leu Glu Lys Glu Val Glu Lys Lys Leu Glu Ser Lys
625 630 635 640

Ser Gly Asn Lys Asn Lys Met Glu Ala Lys Ala Gln Ala Asn Ser Gln
645 650 655

Lys Asp Glu Ile Phe Ala Leu Ile Asn Lys Glu Ala Asn Arg Asp Ala
660 665 670

Arg Ala Ile Ala Tyr Thr Gln Asn Leu Lys Gly Ile Lys Arg Glu Leu
675 680 685

Ser Asp Lys Leu Glu Lys Ile Ser Lys Asp Leu Lys Asp Phe Ser Lys
690 695 700

Ser Phe Asp Glu Phe Lys Asn Gly Lys Asn Lys Asp Phe Ser Lys Ala
705 710 715 720

Glu Glu Thr Leu Lys Ala Leu Lys Gly Ser Val Lys Asp Leu Gly Ile
725 730 735

Asn Pro Glu Trp Ile Ser Lys Val Glu Asn Leu Asn Ala Ala Leu Asn
740 745 750

Glu Phe Lys Asn Gly Lys Asn Lys Asp Phe Ser Lys Val Thr Gln Ala
755 760 765

Lys Ser Asp Leu Glu Asn Ser Val Lys Asp Val Ile Ile Asn Gln Lys
770 775 780

Val Thr Asp Lys Val Asp Asn Leu Asn Gln Ala Val Ser Val Ala Lys
785 790 795 800

Ala Met Gly Asp Phe Ser Arg Val Glu Gln Val Leu Ala Asp Leu Lys
805 810 815

Asn Phe Ser Lys Glu Gln Leu Ala Gln Gln Ala Gln Lys Asn Glu Asp
820 825 830

Phe Asn Thr Gly Lys Asn Ser Glu Leu Tyr Gln Ser Val Lys Asn Ser
835 840 845

Val Asn Lys Thr Leu Val Gly Asn Gly Leu Ser Gly Ile Glu Ala Thr

850

855

860

Ala Leu Ala Lys Asn Phe Ser Asp Ile Lys Lys Glu Leu Asn Glu Lys
865 870 875 880

Phe Lys Asn Phe Asn Asn Asn Asn Gly Leu Lys Asn Ser Thr Glu
885 890 895

Pro Ile Tyr Ala Lys Val Asn Lys Lys Thr Gly Gln Val Ala Ser
900 905 910

Pro Glu Glu Pro Ile Tyr Thr Gln Val Ala Lys Lys Val Asn Ala Lys
915 920 925

Ile Asp Arg Leu Asn Gln Ile Ala Ser Gly Leu Gly Gly Val Gly Gln
930 935 940

Ala Ala Gly Phe Pro Leu Lys Arg His Asp Lys Val Asp Asp Leu Ser
945 950 955 960

Lys Val Gly Leu Ser Ala Ser Pro Glu Pro Ile Tyr Ala Thr Ile Asp
965 970 975

Asp Leu Gly Gly Pro Phe Pro Leu Lys Arg His Asp Lys Val Asp Asp
980 985 990

Leu Ser Lys Val Gly Arg Ser Arg Asn Gln Glu Leu Ala Gln Lys Ile
995 1000 1005

Asp Asn Leu Asn Gln Ala Val Ser Glu Ala Lys Ala Gly Phe Phe
1010 1015 1020

Gly Asn Leu Glu Gln Thr Ile Asp Lys Leu Lys Asp Ser Thr Lys
1025 1030 1035

Lys Asn Val Met Asn Leu Tyr Val Glu Ser Ala Lys Lys Val Pro
1040 1045 1050

Ala Ser Leu Ser Ala Lys Leu Asp Asn Tyr Ala Ile Asn Ser His
1055 1060 1065

Thr Arg Ile Asn Ser Asn Ile Gln Asn Gly Ala Ile Asn Glu Lys
1070 1075 1080

Ala Thr Gly Met Leu Thr Gln Lys Asn Pro Glu Trp Leu Lys Leu
1085 1090 1095

Val Asn Asp Lys Ile Val Ala His Asn Val Gly Ser Val Ser Leu
1100 1105 1110

Ser Glu Tyr Asp Lys Ile Gly Phe Asn Gln Lys Asn Met Lys Asp
1115 1120 1125

Tyr Ser Asp Ser Phe Lys Phe Ser Thr Lys Leu Asn Asn Ala Val
1130 1135 1140

Lys Asp Ile Lys Ser Gly Phe Thr His Phe Leu Ala Asn Ala Phe
1145 1150 1155

Ser Thr Gly Tyr Tyr Cys Leu Ala Arg Glu Asn Ala Glu His Gly
1160 1165 1170

Ile Lys Asn Val Asn Thr Lys Gly Gly Phe Gln Lys Ser
1175 1180 1185

<210> 9
<211> 201
<212> PRT
<213> Homo sapiens

<400> 9

Ser Ser Gly Pro Ser Ser Ser Leu Asp Asn Gly Asn Ser Leu Asp Val
1 5 10 15

Leu Lys Asn His Val Leu Asn Glu Leu Ile Gln Thr Glu Arg Val Tyr
20 25 30

Val Arg Glu Leu Tyr Thr Val Leu Leu Gly Tyr Arg Ala Glu Met Asp
35 40 45

Asn Pro Glu Met Phe Asp Leu Met Pro Pro Leu Leu Arg Asn Lys Lys
50 55 60

Asp Ile Leu Phe Gly Asn Met Ala Glu Ile Tyr Glu Phe His Asn Asp
65 70 75 80

Ile Phe Leu Ser Ser Leu Glu Asn Cys Ala His Ala Pro Glu Arg Val
85 90 95

Gly Pro Cys Phe Leu Glu Arg Lys Asp Asp Phe Gln Met Tyr Ala Lys
100 105 110

Tyr Cys Gln Asn Lys Pro Arg Ser Glu Thr Ile Trp Arg Lys Tyr Ser
115 120 125

Glu Cys Ala Phe Phe Gln Glu Cys Gln Arg Lys Leu Lys His Arg Leu
130 135 140

Arg Leu Asp Ser Tyr Leu Leu Lys Pro Val Gln Arg Ile Thr Lys Tyr
145 150 155 160

Gln Leu Leu Leu Lys Glu Leu Leu Lys Tyr Ser Lys Asp Cys Glu Gly
165 170 175

Ser Ala Leu Leu Lys Lys Ala Leu Asp Ala Met Leu Asp Leu Leu Lys
180 185 190

Ser Val Asn Asp Ser Met His Gln Ile
195 200

